



Optimisation of Callus Induction in the Leaf and Stem Tissues of the Orange Flesh Sweet potato Variety Bhu Sona

Sweet potato (*Ipomoea batatas* (L.) Lam), belonging to the family Convolvulaceae, is one of the important food crops cultivated globally ~7.4 million hectares, with a total production of ~92 million tons (FAOSTAT, 2019). It is cultivated throughout the tropical and subtropical regions of the world, including India (Edison et al., 2009; Tava and Nedunchezhiyan, 2012). Because of its short growing period, ability to grow in diverse environmental conditions and high yield potential, sweet potato is considered as one of the potential crops that can help us to meet the future energy and nutritional needs of both human and livestock populations particularly in developing countries (Motsa et al., 2015; de Albuquerque et al., 2019). The introduction of β -carotene rich orange-fleshed sweet potato varieties in dietary programs has alleviated vitamin-A deficiency among children and pregnant women in many developing countries (Girard et al., 2017; Govender et al., 2019). In India, ICAR-CTCRI and different AICRP-TC centres have released six β -carotene rich varieties viz., Bhu Sona, Bhu Kanti, BhuJa, Gouri, Kamala Sundari and Co-5 (Sunitha et al., 2018). Development of *in vitro* plant regeneration from various plant tissues is important for various basic biotechnology applications such as virus elimination, germplasm conservation, including plant propagation and genetic improvement (Arathi et al., 2019; Ravi et al., 2020; Lenka et al., 2018). In tissue culture, plant hormones influence the differentiation and growth of the explants (Hill and Schaller, 2013). In addition to these, several factors, including genetic factors/genotype differences, the type of explants used for regeneration, the selection of hormones and the concentration used, are also known to determine the regeneration efficiency of the plants (Salari et al., 2013).

Several studies have shown that the callus induction and regeneration efficiency in many crop plants, including sweet potato is highly genotype dependent (Ravi and Indira, 1999; Salari et al., 2013; Arathi et al., 2019). Due to these differential responses of the cultivars, genotype specific callus induction protocol needs to be developed for *in vitro* culture based genetic improvement studies (Salari et al., 2013; Arathi et al., 2019). Moreover, development of callus induction protocols would be helpful in the improvement of important traits through

biotechnological approaches (Muthusamy et al., 2017; Lenka et al., 2019; Jagannadham et al., 2021). Arathi et al., (2019) used the growth regulators viz., BAP, TDZ, GA₃ and IBA and found that the MS media supplemented with 0.5 mg l⁻¹ BAP and 1.0 mg l⁻¹ GA₃ displayed good shooting efficiency in sweet potato. Several workers had shown that Naphthalene Acetic Acid (NAA), a synthetic auxin hormone has good effect on callus induction in many crops (Ahmad and Spoor, 1999; Nazir et al., 2020). The effect of NAA on induction of callusing is yet to be studied in the orange-fleshed sweet potato Bhu Sona. Hence, in the present study, the NAA hormone was used for optimisation of callus induction in leaf, stems and root tissues of the Bhu Sona variety.

The study was conducted in the ICAR-Central Tuber Crops Research Institute (CTCRI), Thiruvananthapuram, during the period 2020-2021. Bhu Sona, sweet potato variety of ICAR-CTCRI, was used in this study to optimise callus induction. Microbial contamination emanating from both endogenous and exogenous sources remains one of the serious concerns in the *in vitro* culture (Amissah et al., 2016). To reduce microbial contamination from the outside environment, first, we initiated the *in vitro* cultures for multiplication of the variety Bhu Sona. The four weeks old plantlets were cut into a smaller size of 3-4 cm shoots and multiplied in the sterile tubes containing MS media with 3% sucrose and 0.8% charcoal. The plantlets were grown in controlled conditions at 25±2°C temperature under a photoperiod of 16 h light and 8-h dark in the plant tissue culture room. Contamination free, four weeks old grown plantlets with uniform size were selected to obtain explants required for callus induction study (Fig. 1).



Fig. 1. Initiation of contamination free fresh *in vitro* plantlets of Bhu Sona.

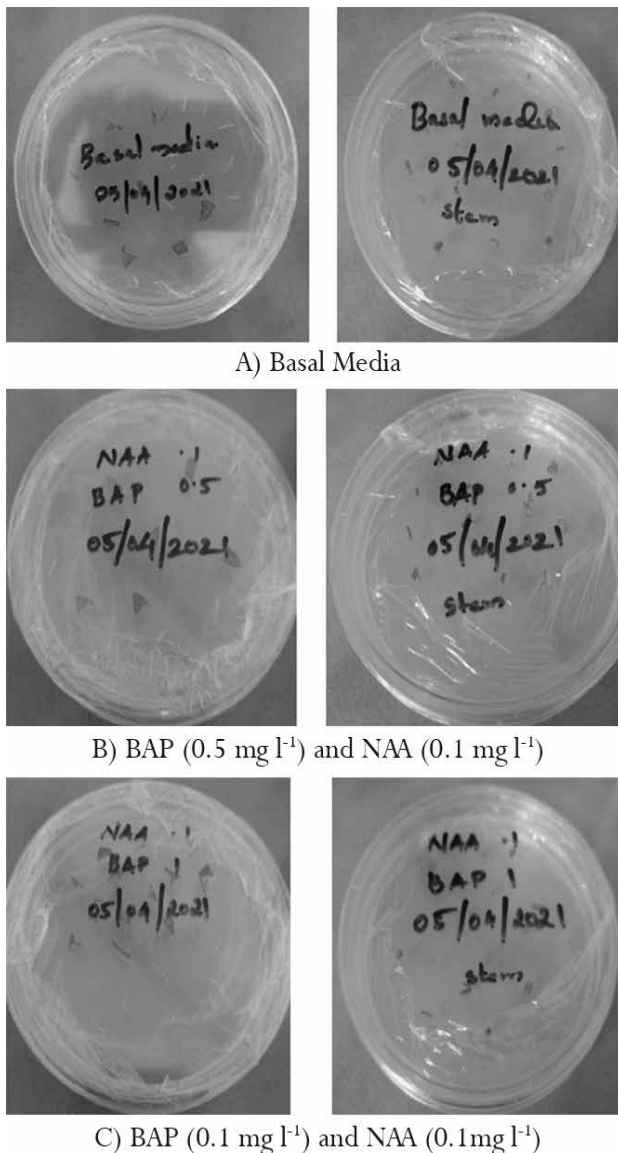


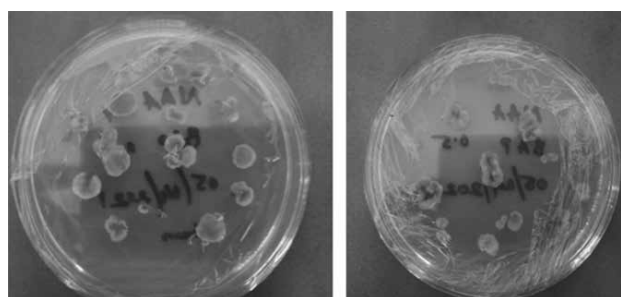
Fig. 2. Culturing of leaf, stem and root tissues of sweet potato for induction of callusing in MS media with different hormone combinations.

The callus induction potential of the variety Bhu Sona was studied in different hormonal combinations viz., Basal media (MS media supplemented with 3% sucrose); basal media supplemented with 0.1 mg l⁻¹ NAA+0.1 mg l⁻¹ BAP and basal media supplemented with 0.1 mg l⁻¹ NAA+0.5 mg l⁻¹ BAP (Table. 1 and Fig. 2). Leaf, stem and root segments, about (0.3 to 0.8 cm) long were excised from four weeks-old *in vitro* raised plants and cultured in the petri dishes containing three media combinations viz., Basal media (MS media with 3% sucrose), basal media supplemented with 0.1 mg l⁻¹ NAA+0.1 mg l⁻¹ BAP and basal media supplemented with 0.1 mg l⁻¹ NAA+0.5 mg l⁻¹ BAP were used to study the effects of plant growth regulators on callus induction from leaf, stem and root tissue of sweet potato. For evaluating callusing efficiency of the leaf tissues 14, 16, and 14 leaf explants were placed in basal media, 0.1 mg l⁻¹ NAA+0.5 mg l⁻¹ BAP and 0.1 mg l⁻¹ NAA+0.1 mg l⁻¹ BAP MS plates, respectively whereas, for evaluating stem tissues 18, 13 and 12 stem explants were placed in basal media, 0.1 mg l⁻¹ NAA+0.5 mg l⁻¹ BAP and 0.1 mg l⁻¹ NAA+0.1 mg l⁻¹ BAP MS plates, respectively (Table 1 and Fig. 2) and for evaluating root tissue, 8, 6 and 8 root explants were placed in 0.1 mg l⁻¹ NAA 0.5 mg l⁻¹ BAP and 0.1 mg l⁻¹ NAA+0.1 mg l⁻¹ BAP MS plates, respectively (Table 1 and Fig. 2). The petri plates were then sealed with parafilm and cultures were incubated in a culture room at 25±2°C temperature under a dark condition. Additionally, charcoal (0.8%) was included in the growth media to act as an adsorbent of phenolic compounds of the cultures (Arathi et al., 2019).

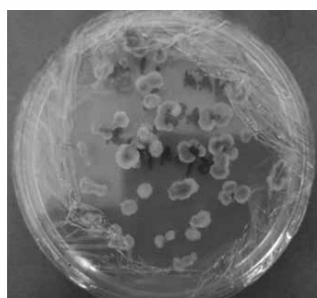
The plates were incubated in dark conditions with a room temperature of 25±2°C in the tissue culture room. One hundred percent callus induction efficiency was observed in both leaf and stem tissues on the MS media supplemented with the hormones NAA and BAP, whereas, in basal media lacking these hormones failed to show the callus induction (Table 1 and Fig. 3). However, the root tissues failed to show callus induction in all three

Table 1. Callus induction in leaf, stem and root tissues of sweet potato under different hormone treatments

Explant	Media Composition	No of Explants inoculated	Callus induced
Leaf	Basal media	14	0
	MS+0.1 mg l ⁻¹ NAA+0.5 mg l ⁻¹ BAP	16	16
	MS+0.1 mg l ⁻¹ NAA+1 mg l ⁻¹ BAP	14	14
Stem	Basal media	18	0
	MS+0.1 mg l ⁻¹ NAA+0.5 mg l ⁻¹ BAP	13	13
	MS+0.1 mg l ⁻¹ NAA+1 mg l ⁻¹ BAP	12	12
Root	Basal media	8	0
	MS+0.1 mg l ⁻¹ NAA+0.5 mg l ⁻¹ BAP	6	0
	MS+0.1 mg l ⁻¹ NAA+1 mg l ⁻¹ BAP	8	0



A) BAP (0.5 mg l^{-1}) and NAA (0.1 mg l^{-1})
(leaf and stem tissues)



B) BAP (0.1 mg l^{-1}) and
(0.1 mg l^{-1}) (leaf and
stem tissues)

Fig. 3. Callusing in leaf, stem and root tissues
of sweet potato

media compositions (Table 1 and Fig. 3). A combination of NAA and BAP has shown success in the induction of callusing in many plants (Ahmad and Spoor, 1999; Nazir et al., 2020). Arathi et al. (2019) have used the growth regulators viz., BAP and IBA for induction of the callusing in sweet potato varieties Gowri and Bhu Sona, however, the callus developed from the leaf and stem tissues failed in organogenesis. Hence, in this study we have used the growth regulators viz., BAP and NAA for callus induction. However, the callus developed from the leaf and stem tissues were failed in organogenesis. Thus, further studies with different hormone combinations, including statistical parameters would be helpful in the standardisation of robust protocol for callus induction and regeneration of the explants in sweet potato.

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References

- Ahmad, S. and Spoor W., 1999. Effects of NAA and BAP on Callus Culture and Plant Regeneration in Curly Kale (*Brassica oleraces* L.). *Pak. J. Biol. Sci.*, **2**:109-112.
- Amissah, S., Coleman, P.A., Sintim, H.Y. and Akromah, R. 2016. In vitro control of microbial contamination of sweet potatoes cultured with nodal explants. *Annu. Res. Rev. Biol.*, **9**(3):1-8. <https://doi.org/10.9734/ARRB/2016/22995>.
- Arathi, S., Hegde, V., Sailekshmi, N. and Koundinya, A.V.V. 2019. An efficient micropropagation protocol for nutritionally rich varieties of sweet potato (*Ipomoea batatas* L.). *J. Root Crops*, **45**(2):12-18.
- de Albuquerque, T.M.R., Sampaio, K.B. and de Souza, E.L. 2019. Sweet potato roots: Unrevealing an old food as a source of health promoting bioactive compounds—A review. *Trends Food Sci. Technol.*, **85**:277-286.
- Edison, S., Hegde, V., Makesh Kumar, T., Srinivas, T., Suja, G., Padmaja, G. 2009. Sweet potato in the Indian Sub-Continent. In: Loebenstein, G., Thottappilly, G. (eds) *The Sweet potato*. Springer, Dordrecht, pp. 391-414.
- FAOSTAT (2019). Food Agriculture and Organization (FAOSTAT). Retrieved from <https://www.fao.org/faostat/en/#home>
- Girard, A.W., Grant, F., Watkinson, M., Okuku, H.S., Wanjala, R., Cole, D., Levin, C. and Low, J. 2017. Promotion of orange-fleshed sweet potato increased vitamin A intakes and reduced the odds of low retinol-binding protein among postpartum Kenyan women. *J. Nutr.*, **147**(5):955-963.
- Govender, L., Pillay, K., Siwela, M., Modi, A.T. and Mabhaudhi, T. 2019. Improving the dietary vitamin A content of rural communities in South Africa by replacing non-biofortified white maize and sweet potato with biofortified maize and sweet potato in traditional dishes. *Nutrients*, **11**(6):1198. <https://doi.org/10.3390/nu11061198>.
- Hill, K. and Schaller, G.E. 2013. Enhancing plant regeneration in tissue culture: a molecular approach through manipulation of cytokinin sensitivity. *Plant Signal Behav.*, **8**(10):212-24.
- Jagannadham, P.T.K., Savadi, S. and Muthusamy, S.K. 2021. Plant genome editing in basic research to understand molecular functions. In: Gupta, O. P. and Karkute, S. G. (eds) *Genome Editing in Plants*. CRC Press, pp. 187-202.
- Lenka, S.K., Muthusamy, S.K., Chinnusamy, V. and Bansal, K.C. 2018. Ectopic expression of rice *PYL3* enhances cold and drought tolerance in *Arabidopsis thaliana*. *Mol. Biotechnol.*, **60**(5):350-361.
- Lenka, S.K., Singh, A.K., Muthusamy, S.K., Smita, S., Chinnusamy, V. and Bansal, K.C., 2019. Heterologous expression of rice RNA-binding glycine-rich (RBG) gene *OsRBGD3* in transgenic *Arabidopsis thaliana* confers cold stress tolerance. *Funct. Plant Biol.*, **46**(5):482-491.
- Motsa, N.M., Modi, A.T. and Mabhaudhi, T. 2015. Sweet potato (*Ipomoea batatas* L.) as a drought tolerant and food security crop. *South African J. Sci.*, **111**(11-12):1-8.
- Muthusamy, S.K., Sivalingam, P.N., Sridhar, J., Singh, D., Haldhar, S.M. and Kaushal, P., 2017. Biotic stress inducible promoters in crop plants—a review. *J. Agri. Ecol.*, **4**:14-24.
- Nazir, S., Jan, H., Tungmunnithum, D., Drouet, S., Zia, M., Hano, C. and Abbasi, B.H. 2020. Callus culture of Thai basil is an effective biological system for the production of antioxidants. *Molecules*, **25**(20):4859. <https://doi.org/10.3390/molecules25204859>
- Ravi, V. and Indira, P. 1999. Crop physiology of sweet potato, Jules Janick (eds.) *Hort. Rev.*, **23**:312-313.

- Ravi, V., Muthusamy, S.K., Raju, S. and Chakrabarti, S.K., 2020. Analysis of molecular chaperones in differentiating storage root compared to non-tuber forming fibrous root of sweet potato (*Ipomoea batatas*). *Current Horticulture*, **8**(2):51-56.
- Salari, A., Sharma, A., Muthusamy, S.K., Singh, S.K., Chinnusamy, V. and Bansal, K.C., 2013. An improved protocol for high frequency plant regeneration from mature embryos of wheat. *Proc. Indian Natl. Sci. Acad.*, **79**:159-166.
- Sunitha, S., George, J., Sheela, M.N., Sureshkumar, J. and Mukherjee, A. 2018. Tuber Crops varieties released recommended for release by AICRP on tuber crops over five decades, Technical Bulletin No. 71, ICAR-All India Coordinated Research Project on Tuber Crops, ICAR-CTCRI, Thiruvananthapuram, Kerala, India, 140 p
- Tavva, S. and Nedunchezhiyan, M., 2012. Global status of sweet potato cultivation. *Fruit Veg. Cereal Sci. Biotechnol*, **6**:143-147.

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